



# Chiral discrimination of enantiomers with a self-assembled monolayer of functionalized $\beta$ -cyclodextrins on Au surfaces

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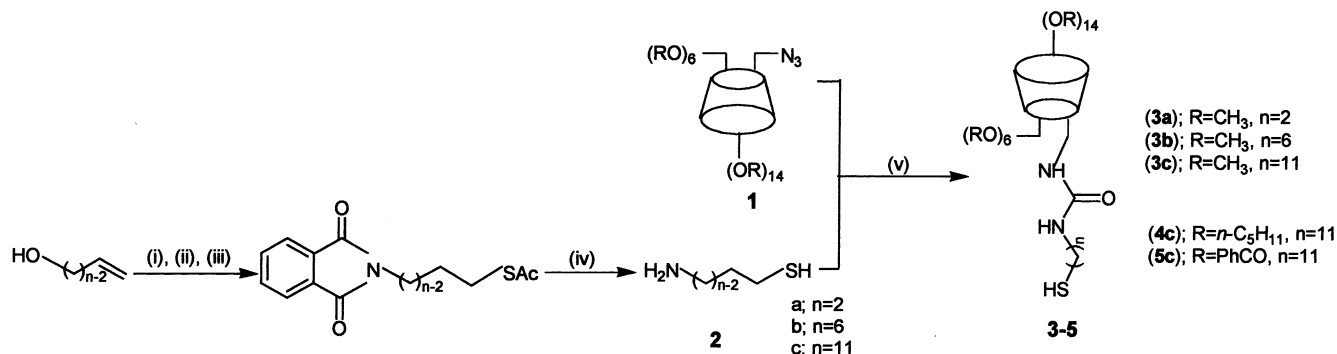
**Abstract**—A facile synthetic route to mercaptyl functionalized  $\beta$ -cyclodextrins was devised and self-assembled onto the gold electrodes of a quartz crystal microbalance (QCM) to afford a durable sensor capable of chiral discrimination of enantiomers. © 2002 Published by Elsevier Science Ltd.

Molecular host–guest recognition systems have attracted significant attention in recent years.<sup>1</sup> It becomes evident that researchers in this area are increasingly interested in the potential applications of these systems in chemical sensors and molecular electronics.<sup>2</sup> Among synthetic host molecules, cyclodextrin (CD) has emerged as an ideal candidate for this pursuit due to its well-defined molecular cavity and its ability to accommodate a variety of guest molecules.<sup>3</sup> It is necessary for cyclodextrin to be confined or immobilized on the surface of a substrate such as the Au surface at the device level. Therefore, several routes for the synthesis of dialkyl sulfide- and alkanethiol-substituted cyclodextrins forming self-assembled monolayers (SAMs) on Au and Ag have been reported. Rojas et al.<sup>1</sup> immobilized native  $\beta$ -CD molecules on a gold surface through thiol moieties directly linked to the  $\beta$ -CD molecule. Nelles et al.<sup>4</sup> reported the immobilization

kinetics of  $\beta$ -CD through mono- or multithiol linkers. More recently, Beulen et al.<sup>5</sup> developed synthetic routes to introduce short and long dialkyl sulfides onto the primary side of  $\alpha$ -,  $\beta$ -, and  $\gamma$ -CDs. However, it is apparent that these synthetic routes were invariably applied to native CDs whilst a systematic approach towards dialkyl sulfide- or thiol-modified functionalized CDs was not reported.

The present work reports on a facile synthetic approach to structurally well-defined mercaptyl functionalized  $\beta$ -CDs. Subsequently, self assembly onto an Au surface of a quartz crystal microbalance (QCM) afforded a durable enantioselective coating which was applicable for the chiral discrimination of enantiomers.

Scheme 1 depicts the synthetic route to mercaptyl functionalized  $\beta$ -CD from (6<sup>A</sup>-azido-6<sup>A</sup>-deoxy)heptakis(2,3-



**Scheme 1.** Reagents and conditions: (i) imidazole/Ph<sub>3</sub>P/I<sub>2</sub>/DCM (dry); (ii) potassium phthalimide/DMF (dry)/ $\Delta$ ; (iii) CH<sub>3</sub>COSH/hexane (dry)/UV; (iv) NH<sub>2</sub>NH<sub>2</sub>·H<sub>2</sub>O/MeOH/reflux; (v) PPh<sub>3</sub>/CO<sub>2</sub>/THF (dry).

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di-*O*-functionalized)-6<sup>B</sup>,6<sup>C</sup>,6<sup>D</sup>,6<sup>E</sup>,6<sup>F</sup>,6<sup>G</sup>-hexa-*O*-functionalized- $\beta$ -CD (**1**). The synthesis of (**1**) has been reported previously,<sup>6</sup> thus the crucial part of the whole scheme relies on the availability of mercaptyl-alkylamines of different lengths, NH<sub>2</sub>(CH<sub>2</sub>)<sub>*n*</sub>SH (**2**). With *n*=2, commercially available cysteamine was used directly. For *n*>2, a facile route was derived for the synthesis of 6-( $\omega$ -mercaptyl)-hexanamine (**2b**) and 11-( $\omega$ -mercaptyl)-undecanamine (**2c**) using a modified Gabriel reaction<sup>7</sup> followed by a photoaddition reaction<sup>8</sup> and then a deprotection step which conveniently cleaved both protective groups in one step (step iv). Coupling of (**1**) and the thiol linker (**2**) was accomplished using a Staudinger reaction under mild conditions and the expected products (**3**) were obtained.<sup>9</sup> Table 1 summarizes the respective yields of these mercaptyl  $\beta$ -CD derivatives, in which R=methyl, pentyl, and benzoyl, respectively. The formation of the carbamido moiety in these compounds was evident from the <sup>13</sup>C NMR resonance at ca.  $\delta$  159 ppm.

Immobilization of mercaptyl  $\beta$ -CDs (**3a**)–(**5c**) on the Au surface was achieved using established procedures.<sup>10</sup> Subsequently, XPS studies of the Au surface revealed the presence of the elements C, N, O and S. Moreover, the presence of a broad peak with binding energy at ca. 162 eV for the S<sub>2p</sub> environment is consistent with the presence of an Au–S moiety due to self-assembly of the thiols on the Au surface.<sup>4</sup> Further corroborative evidence of the successful immobilization of these  $\beta$ -CD derivatives on the Au surface was obtained from positive 10 KeV Ar TOF-SIMS spectra (Table 2). The presence of two distinct groups of fragments at molecular mass [*M*+Au]<sup>+</sup> and [*M*+Au–S]<sup>+</sup>, where *M* is the molecular mass of the

**Table 1.** Yields of mercaptyl functionalized persubstituted  $\beta$ -CD derivatives

Compd	<i>n</i>	Yield (%)
<b>2a</b>	2	–
<b>2b</b>	6	82
<b>2c</b>	11	87
<b>3a</b>	2	19
<b>3b</b>	6	25
<b>3c</b>	11	34
<b>4c</b>	11	29
<b>5c</b>	11	30

**Table 2.** Immobilization of mercaptyl functionalized  $\beta$ -CD derivatives on the Au electrodes of a QCM

SAMs	<b>3a</b>	<b>3c</b>	<b>4c</b>	<b>5c</b>
Surface concentration ( $\times 10^{11}$ mol/cm <sup>2</sup> )	6.90	8.98	4.91	3.44
TOF-SIMS	[ <i>M</i> +Au–S] <sup>+</sup> <sup>a</sup>	1807	2931	3609
	[ <i>M</i> +Au] <sup>+</sup>	1713	2963	3641
	[ <i>M</i> +Au <sub>2</sub> ] <sup>+</sup>	1910	3160	3840
$\alpha_{R/S}$	1.114	1.125	1.145	1.178

<sup>a</sup> *m/z*, *M* is the molecular mass of the mercaptyl functionalized  $\beta$ -CD molecule, e.g. *M*<sub>3a</sub>=1517.2 g/mol.

<sup>†</sup> Lab Crystal Oscillator and AT-cut 10 MHz quartz plates were purchased from International Crystal Manufacturer Inc., Oklahoma City, OK. The time-course frequency signals were monitored by TF830 universal frequency counter and recorded by PC.

<sup>‡</sup>  $\alpha_{R/S} = \Delta f_{\max-R} / \Delta f_{\max-S}$ .  $\Delta f_{\max-R}$ ,  $\Delta f_{\max-S}$  standing for the maximum frequency reductions of a QCM while exposed to the respective enantiomeric analyte.<sup>15</sup>

mercaptyl  $\beta$ -CD molecule (i.e. *M*<sub>3a</sub>=1517.2 g/mol) confirms the formation of the Au–S covalent bond between the mercaptyl  $\beta$ -CD molecule and the Au substrate.<sup>11</sup>

A study of the gas phase chiral discrimination of enantiomers by immobilized  $\beta$ -CD monolayers was carried out using an AT-cut 10 MHz quartz crystal microbalance (QCM). Compounds (**3a**)–(**5c**) were immobilized on the surface of the Au electrodes of a QCM, and the surface concentration of the immobilized  $\beta$ -CD host molecules was calculated according to the equation:<sup>12</sup>  $\Delta f = (2.26 \times 10^{-6}) f_0^2 (\Delta m/A)$  and summarized in Table 2.

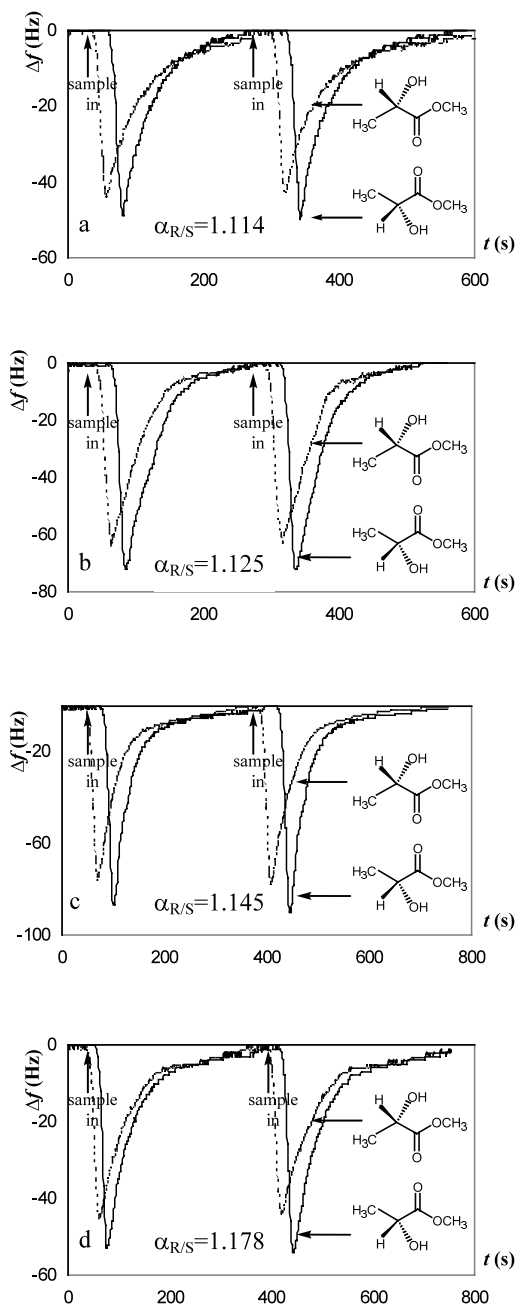
The influence of the thiol linker length (*n*) on the surface concentration of immobilized  $\beta$ -CD monolayers was well reflected in SAM (**3a**) and (**3c**), where the longer thiol linker (*n*=11) affords compound (**3c**) which is more densely packed in the monolayer.

The chiral discrimination experiment was carried out in a home-made pulse measuring system using pre-dried N<sub>2</sub> as carrier gas, at 200 ml/min.<sup>13</sup> The coated QCM sensors were exposed to the same concentration of (+)-methyl lactate and (–)-methyl lactate analytes at 25±0.1°C. Their respective time-course signals were recorded and are plotted in Fig. 1.<sup>†</sup>

As can be seen, the responses of the QCM sensors due to the inclusion complexation of the analyte molecule by the  $\beta$ -CD monolayer are reproducible. Moreover, the responses towards the respective chiral forms of methyl lactate are similar but differ in the extent of frequency reductions ( $\Delta f$ ), reflecting the differences in the binding affinity of the  $\beta$ -CD monolayers towards the respective enantiomer.<sup>14</sup> All four  $\beta$ -CD monolayers depict higher binding affinity towards the (+)-methyl lactate than the (–)-form.

Amongst the  $\beta$ -CD SAMs, (**3c**) depicts better chiral discrimination as measured by the chiral discrimination factor,  $\alpha_{R/S}$ ,<sup>‡</sup> than (**3a**) corresponding to the increase in the surface concentration of the immobilized permethylated  $\beta$ -CD molecules on the Au surface.

The more crowded  $\beta$ -CD cavity in the molecular structures of (**4c**) and (**5c**) probably enhances the non-covalent interactions between the interior cavity and the



**Figure 1.** Chiral discrimination of four QCM sensors towards (+)-methyl lactate and (–)-methyl lactate. (a) **3a**; (b) **3c**; (c) **4c**; (d), **5c**. Temp.  $25 \pm 0.1^\circ\text{C}$ ; carrier gas  $\text{N}_2$ , 200 ml/min; sample injection, 2.0  $\mu\text{l}$ .

included methyl lactate molecules, thus affording improved chiral discrimination:  $\alpha_{R/S-5c} > \alpha_{R/S-4c} > \alpha_{R/S-3c}$  despite the decreased surface concentration of the immobilized derived  $\beta$ -CD molecules.

In summary, Scheme 1 depicts a facile synthetic route for the synthesis of mercaptyl functionalized  $\beta$ -CD derivatives. Analytical data for compounds (**2**)–(**5**) is detailed in Ref. 16. These can be self-assembled onto the Au electrodes of a QCM. The application of these structurally defined  $\beta$ -CD monolayers for the study of gas phase chiral discrimination of enantiomers of

methyl lactate were successful. It was found that the perbenzoylated  $\beta$ -CD monolayer depicted the best enantiomeric discriminatory ability towards methyl lactate. Further investigations on the host–guest interactions between chiral ligands and the monolayers are currently underway.

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## 16. Analytical data for compounds 2–5:

**6-( $\omega$ -Mercapto)-hexanamine (2b):** IR ( $\text{cm}^{-1}$ ) 3382 (N–H str), 2940 (C–H str), 2502 (S–H str), 1595 (–NH<sub>2</sub> def), 1065 (C–NH<sub>2</sub> str); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  (ppm) 2.66 (m, 2H, CH<sub>2</sub>–SH), 2.49 (t,  $J=7.2$  Hz, 2H, CH<sub>2</sub>–NH<sub>2</sub>), 1.67 (m, 2H, CH<sub>2</sub>–CH<sub>2</sub>–NH<sub>2</sub>), 1.41 (m, 6H), 0.96 (t,  $J=7.2$  Hz, 1H, SH); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  (ppm) 41.9 (NH<sub>2</sub>CH<sub>2</sub>–), 38.9, 33.4, 29.0, 28.2 (–(CH<sub>2</sub>)<sub>4</sub>–), 26.3 (–CH<sub>2</sub>–SH); accurate MS  $m/z$  calcd 133.0925, found 133.0921.

**11-( $\omega$ -Mercapto)-undecanilamine (2c):** IR ( $\text{cm}^{-1}$ ) 3372 (N–H str), 2920 (C–H str), 2510 (S–H str), 1595 (–NH<sub>2</sub> def.), 1065 (C–NH<sub>2</sub> str); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  (ppm) 2.66 (t,  $J=7.2$  Hz, 2H, CH<sub>2</sub>–SH), 2.50 (t,  $J=7.2$  Hz, 2H, CH<sub>2</sub>–NH<sub>2</sub>), 1.59 (m, 2H, CH<sub>2</sub>–CH<sub>2</sub>–NH<sub>2</sub>), 1.26–1.40 (m, 17H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  (ppm) 42.0 (NH<sub>2</sub>CH<sub>2</sub>–), 39.1, 33.9, 33.6, 29.3, 29.1, 28.9, 28.4, 28.2, 26.8 (–(CH<sub>2</sub>)<sub>9</sub>–), 24.5 (–CH<sub>2</sub>–SH); accurate MS  $m/z$  calcd 203.1708, found 203.1705.

**6<sup>A</sup>- $\omega$ -Mercapto-ethylureado-6<sup>A</sup>-deoxyheptakis(2,3-di-*O*-methyl)-6<sup>B</sup>,6<sup>C</sup>,6<sup>D</sup>,6<sup>E</sup>,6<sup>F</sup>,6<sup>G</sup>-hexa-*O*-methyl- $\beta$ -cyclodextrin (3a):** IR ( $\text{cm}^{-1}$ ) 3396 (N–H str), 2931 (C–H str), 1668 (C=O str, urea), 1040 (C–O–C str); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  (ppm) 158.72 (–NH–CO–NH–), 98.73 (C–1), 81.66 (C–2), 80.24 (C–3), 80.02 (C–4), 71.16 (C–6b), 70.83 (C–5), 67.73 (C–6a), 61.11 (H<sub>3</sub>CO–C3), 58.81 (H<sub>3</sub>CO–C6), 58.28 (H<sub>3</sub>CO–C2), 40.82 (–NHCH<sub>2</sub>–), 25.43 (–CH<sub>2</sub>–SH); ESI-MS  $m/z$  Calcd 1517.4, found 1517.2; elemental analysis calcd: C, 51.45; H, 7.70; N, 1.84; S, 2.11; found: C, 51.54; H, 7.51; N, 1.93; S, 1.95%.

**(6<sup>A</sup>- $\omega$ -Mercapto-hexanureado-6<sup>A</sup>-deoxy)heptakis(2,3-di-*O*-methyl)-6<sup>B</sup>,6<sup>C</sup>,6<sup>D</sup>,6<sup>E</sup>,6<sup>F</sup>,6<sup>G</sup>-hexa-*O*-methyl- $\beta$ -cyclodextrin (3b):** IR ( $\text{cm}^{-1}$ ) 3399 (N–H str), 2930 (C–H str), 1657 (C=O str, urea), 1037 (C–O–C str); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  (ppm) 159.00 (NH–CO–NH), 98.76 (C–1), 81.66 (C–2), 81.32 (C–3), 80.22 (C–4), 71.40 (C–6b), 70.84 (C–5), 67.54 (C–6a), 61.18 (H<sub>3</sub>CO–C3), 58.90 (H<sub>3</sub>CO–C6), 58.47 (H<sub>3</sub>CO–C2), 41.22 (–NHCH<sub>2</sub>–), 38.67, 30.07, 28.99, 28.14 (–(CH<sub>2</sub>)<sub>4</sub>–), 26.44 (–CH<sub>2</sub>–SH); ESI-MS  $m/z$  calcd 1573.5; found: 1596.5 for [M+Na]<sup>+</sup>; elemental analysis calcd C,

52.67; H, 7.94; N, 1.78; S, 2.03; found: C, 52.71; H, 7.79; N, 1.76; S, 1.92%.

**(6<sup>A</sup>- $\omega$ -Mercapto-undecanylureado-6<sup>A</sup>-deoxy)heptakis(2,3-di-*O*-methyl)-6<sup>B</sup>,6<sup>C</sup>,6<sup>D</sup>,6<sup>E</sup>,6<sup>F</sup>,6<sup>G</sup>-hexa-*O*-methyl- $\beta$ -cyclodextrin (3c):** IR ( $\text{cm}^{-1}$ ) 3396 (N–H str), 2928 (C–H str), 1659 (C=O str, urea), 1039 (C–O–C str). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  (ppm) 159.02 (NH–CO–NH), 98.70 (C–1), 81.64 (C–2), 81.35 (C–3), 80.26 (C–4), 71.85 (C–6b), 70.97 (C–5), 67.68 (C–6a), 61.14 (H<sub>3</sub>CO–C3), 58.83 (H<sub>3</sub>CO–C6), 58.36 (H<sub>3</sub>CO–C2), 41.27 (–NHCH<sub>2</sub>–), 40.36–26.80 (–(CH<sub>2</sub>)<sub>9</sub>–), 24.44 (–CH<sub>2</sub>–SH); ESI-MS  $m/z$  calcd 1643.6, found 1666.1 for [M+Na]<sup>+</sup>; elemental analysis calcd C, 54.05; H, 8.22; N, 1.70; S, 1.95; found: C, 54.4; H, 7.82; N, 1.29%; S, 1.70%.

**(6<sup>A</sup>- $\omega$ -Mercapto-undecanylureado-6<sup>A</sup>-deoxy)heptakis(2,3-di-*O*-pentyl)-6<sup>B</sup>,6<sup>C</sup>,6<sup>D</sup>,6<sup>E</sup>,6<sup>F</sup>,6<sup>G</sup>-hexa-*O*-pentyl- $\beta$ -cyclodextrin (4c):** IR ( $\text{cm}^{-1}$ ) 3409 (N–H str), 2933 (C–H str), 1670 (C=O str, urea), 1043 (C–O–C str); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  (ppm) 159.03 (NH–CO–NH), 98.76 (C–1), 81.39 (C–2), 81.19 (C–3), 78.94 (C–4), 74.94 (C–5), 71.79 (C–6b), 67.48 (C–6a), 72.42–72.18 (C–1'), 30.78–29.93 (C–2'), 29.31–29.06 (C–3'), 23.67–23.42 (C–4'), 14.87 (C–5'), 39.64 (–NHCH<sub>2</sub>–), 35.51–24.38 (–(CH<sub>2</sub>)<sub>9</sub>–), 21.50 (–CH<sub>2</sub>–SH); ESI-MS  $m/z$  calcd 2788.7, found 2789.1; elemental analysis calcd C, 66.82; H, 10.82; N, 0.72; S, 1.16; found: C, 66.49; H, 11.21; N, 1.17; S, 1.02%.

**(6<sup>A</sup>- $\omega$ -Mercapto-undecanylureado-6<sup>A</sup>-deoxy)heptakis(2,3-di-*O*-benzoyl)-6<sup>B</sup>,6<sup>C</sup>,6<sup>D</sup>,6<sup>E</sup>,6<sup>F</sup>,6<sup>G</sup>-hexa-*O*-benzoyl- $\beta$ -cyclodextrin (5c):** IR ( $\text{cm}^{-1}$ ) 3412 (N–H str), 3069 (Ar–H str), 2922 (C–H str), 1732 (C=O str, urea), 1610, 1449 (Ar–ring), 1043 (C–O str); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  (ppm) 166.15, 165.88, 164.82 (Ar–CO–), 158.47 (NH–CO–NH), 127.58, 127.72, 129.43, 129.61, 129.97, 132.16, 132.50 (H-bearing aromatic carbon atoms), 128.21, 128.50, 128.64 (quaternary aromatic carbon atoms) 97.57 (C–1), 77.4 (C–4), 71.33 (C–2 and C–3), 69.99 (C–5), 63.49 (C–6b), 63.39 (C–6a), 40.63 (–NHCH<sub>2</sub>–), 34.01–26.94 (–(CH<sub>2</sub>)<sub>9</sub>–), 24.57 (–CH<sub>2</sub>–SH); ESI-MS  $m/z$  calcd 3445.2, found 3468.1 for (M+Na)<sup>+</sup>; elemental analysis calcd: C, 67.61; H, 5.09; N, 0.81; S, 0.93; found: C, 68.04; H, 4.88; N, 0.74; S, 0.82%.